

Using metagenomic sequencing in clinical practice

Natasha Spottiswoode, MD DPhil

Assistant Professor

Division of Infectious Diseases

University of California San Francisco

Disclaimers

- These slides represent consensus views at UCSF; this is an area of evolving evidence
- No personal disclosures

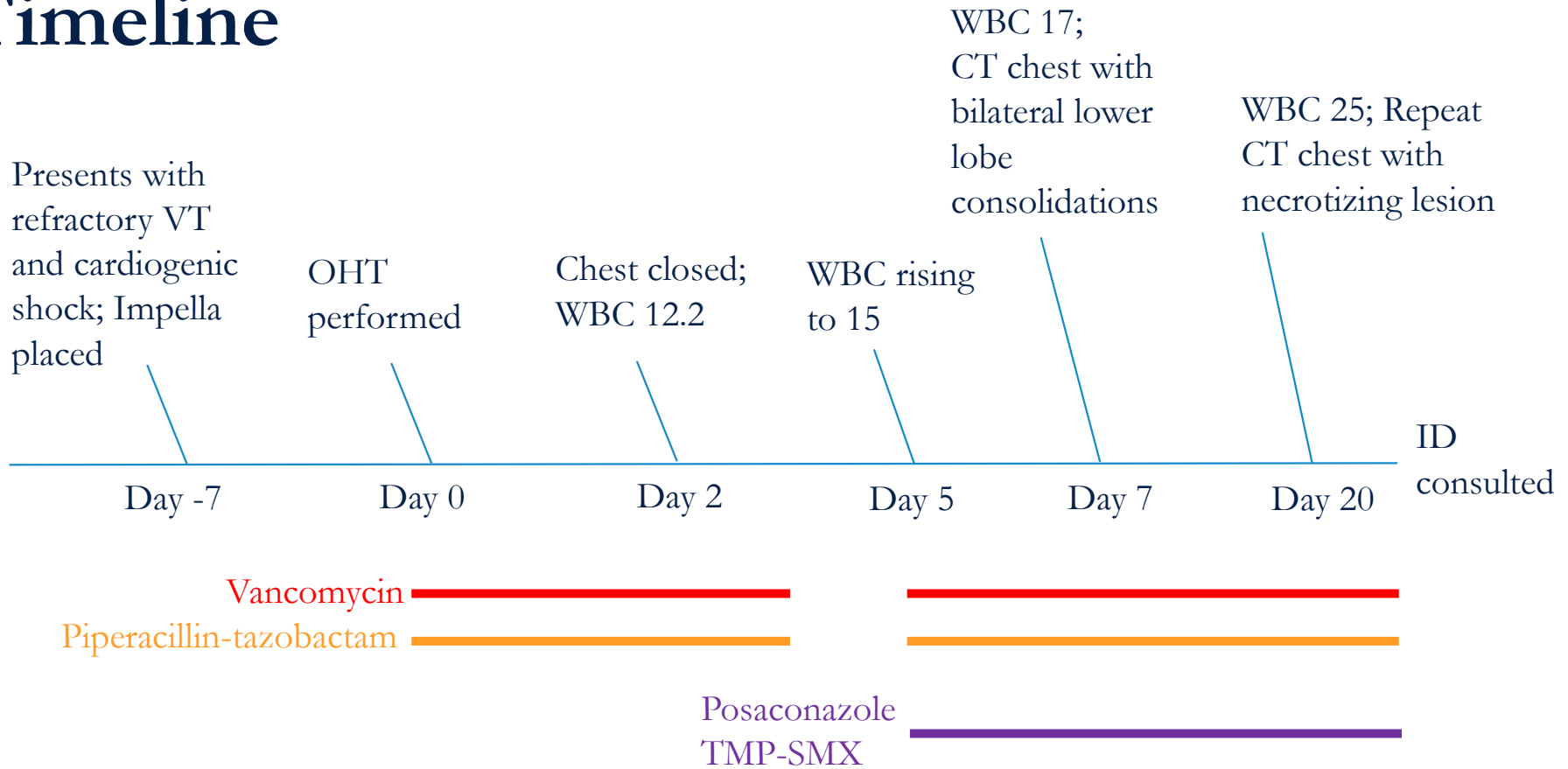


Natasha.spottiswoode@ucsf.edu
[@NSpottiswoodeMD](https://www.instagram.com/NSpottiswoodeMD)

A challenging clinical case

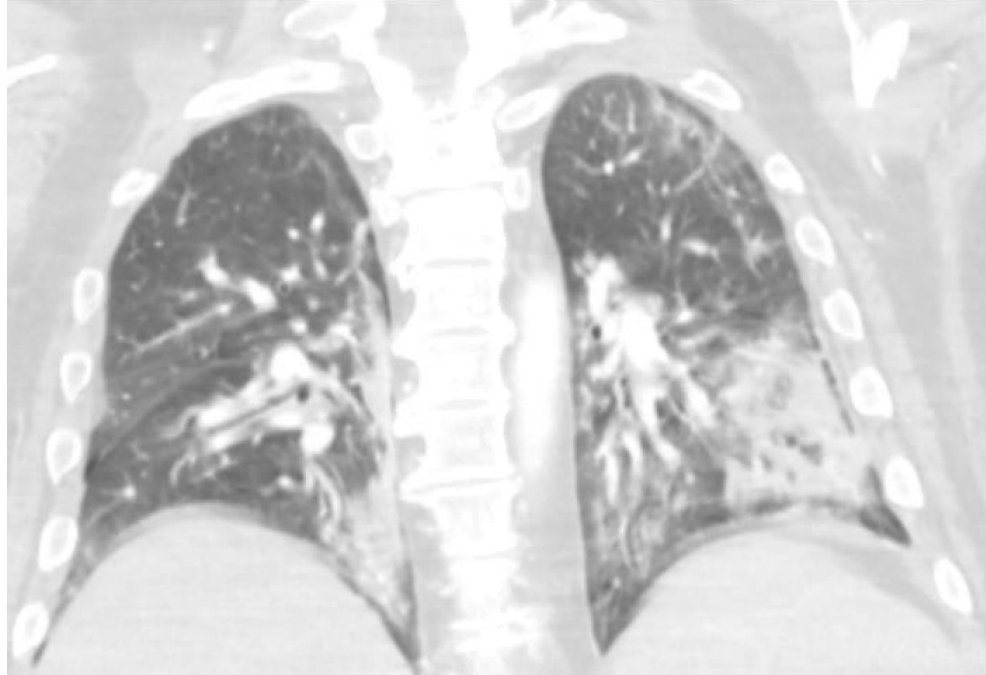
- 56yo with orthotopic heart transplant 3 weeks prior (CMV D+/R+), thymoglobulin induced, on tacrolimus, mycophenolate mofetil and prednisone.
- **Consult question: diagnostics and therapeutics for cavitating pneumonia**

Timeline

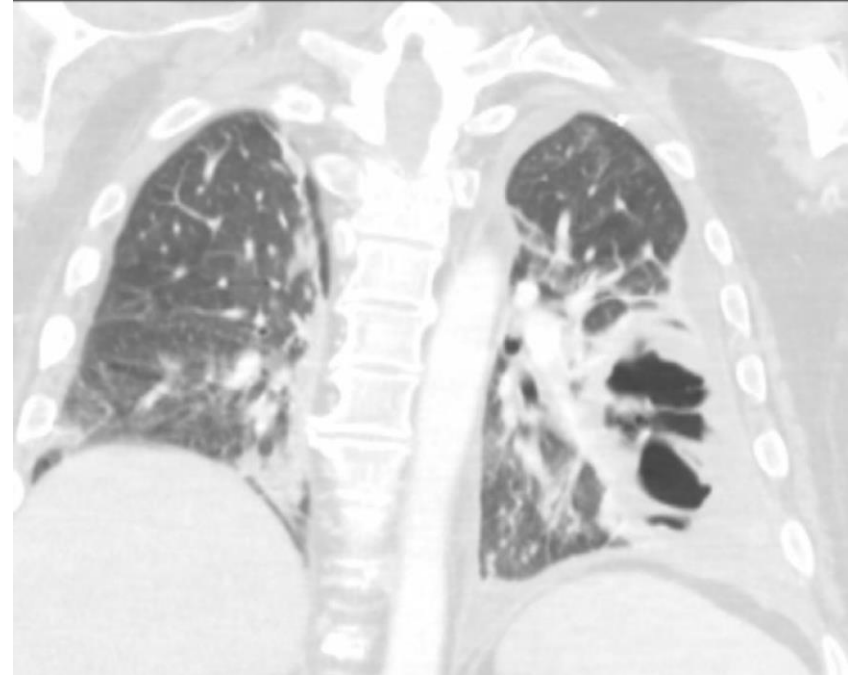


The imaging

Day 7 post-transplant



Day 20 post-transplant



Workup to date

Cultures:

Blood cultures x2 negative

Sputum cultures:

Bacterial with oral flora

Mycobacterial negative

Fungal negative

Antibodies:

Coccidioides antibodies: negative

Histoplasma antibodies: negative

HIV antibody: negative

Quantiferon gold: negative

Antigens:

Cryptococcus Ag negative

Coccidioides Ag: negative

Histoplasma Ag: negative

PCR tests:

COVID negative

Respiratory viral panel negative

Nasal legionella: negative

CMV plasma: negative

Fungal markers:

B-d-glucan: 31 (normal)

Galactomannan: 0.2 (normal)

Pleural fluid from thoracentesis

Bacterial cultures: negative

Mycobacterial cultures: negative to date

Fungal cultures: negative to date

**Pulmonology is consulted for bronchoscopy;
defers pending pleural results**

Question 1: What empiric therapies would you recommend now?

- 1) Continue vancomycin and piperacillin-tazobactam only
- 2) Add azithromycin for atypical organism coverage
- 3) Add liposomal amphotericin for additional fungal coverage
- 4) Call back Pulmonology
- 5) Consult a surgical team

A diagnostic test returns...

TEST RESULTS

MICROBIAL CELL-FREE DNA DETECTED

QUANTITY
MOLECULES PER MICROLITER (MPM)¹

ANNOTATIONS
FURTHER INTERPRETATION OF
ANTIMICROBIAL RESISTANCE (AMR) ON PAGE 2

! **Obligate & Opportunistic Pathogens²** Likely to cause disease in humans at any quantity

✦ Fungi ***Rhizopus microsporus*** 16,756
Alert result

! **Commensal Pathogens & DNA Viruses²** Known to be associated with disease but may also represent normal microbiota

✦ Bacteria ***Staphylococcus epidermidis*** 2,636
coagulase-negative staphylococcus

• **AMR marker: *mecA* detected**
Consistent with resistance to methicillin.

Enterococcus faecium 83

• **AMR marker: *vanA* detected**
Consistent with resistance to vancomycin and other glycopeptides (vancomycin-resistant enterococci, VRE)

¹ Molecules Per Microliter = number of DNA fragments present in one microliter of plasma. Visualization of MPM shows quantile of each detected microbe based on 10,000 specimens with positive, quantitative Karius Test results. No quantile is shown if < 20 detections of the microbe were made in the 10,000 specimens or if the microbe is an obligate or opportunistic pathogen. The analytical range of the assay is 10 - 316,000 MPM.

² Based on a review of Carroll KC, Pfaller MA. 2019. Manual of Clinical Microbiology, 12th Edition. ASM Press, Washington, DC and Bennett JE, Dolin R, Blaser MJ. 2019. Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases, 9th Edition. Elsevier, Philadelphia, PA

- Plasma mNGS returns
- Urgently taken to OR by thoracic surgery for LLL lobectomy
- Liposomal amphotericin started in addition to posaconazole

TEST RESULTS

MICROBIAL CELL-FREE DNA DETECTED

QUANTITY
MOLECULES PER MICROLITER (MPM)¹

ANNOTATIONS
FURTHER INTERPRETATION OF
ANTIMICROBIAL RESISTANCE (AMR) ON PAGE 2

! **Obligate & Opportunistic Pathogens²** Likely to cause disease in humans at any quantity

✦ Fungi ***Rhizopus microsporus*** 16,756
Alert result

! **Commensal Pathogens & DNA Viruses²** Known to be associated with disease but may also represent normal microbiota

🦠 Bacteria ***Staphylococcus epidermidis*** 2,636
coagulase-negative staphylococcus

• **AMR marker: *mecA* detected**
Consistent with resistance to methicillin.

Enterococcus faecium 83

• **AMR marker: *vanA* detected**
Consistent with resistance to vancomycin and other glycopeptides (vancomycin-resistant enterococci, VRE)

¹ Molecules Per Microliter = number of DNA fragments present in one microliter of plasma. Visualization of MPM shows quantile of each detected microbe based on 10,000 specimens with positive, quantitative Karius Test results. No quantile is shown if < 20 detections of the microbe were made in the 10,000 specimens or if the microbe is an obligate or opportunistic pathogen. The analytical range of the assay is 10 - 316,000 MPM.

² Based on a review of Carroll KC, Pfaller MA. 2019. Manual of Clinical Microbiology, 12th Edition. ASM Press, Washington, DC and Bennett JE, Dolin R, Blaser MJ. 2019. Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases, 9th Edition. Elsevier, Philadelphia, PA

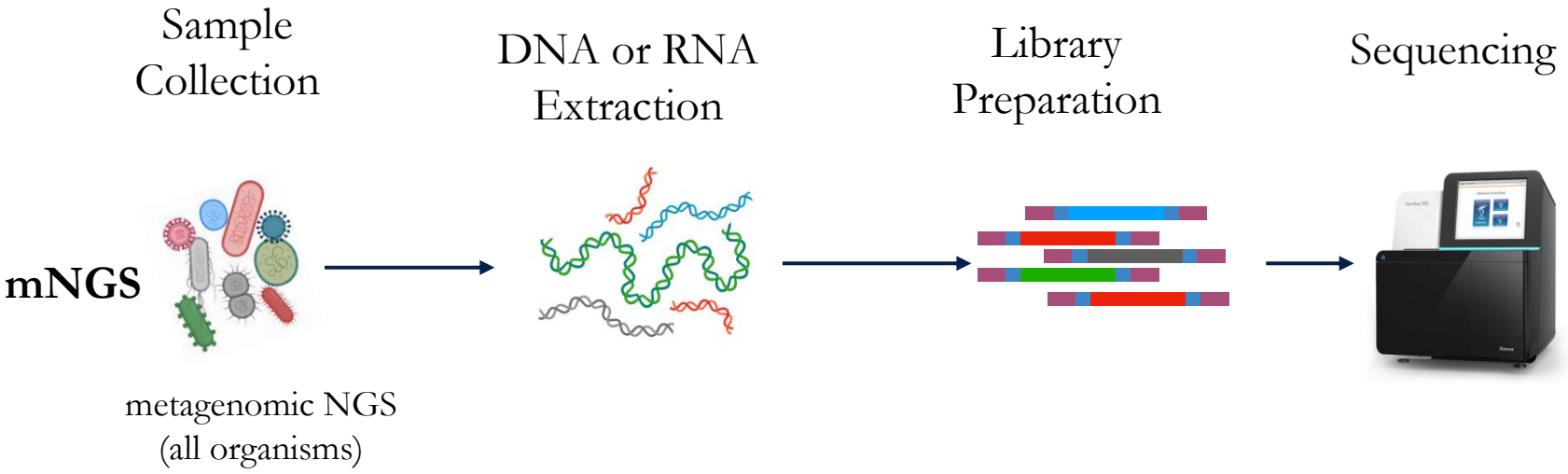
Question 2: With this test result, would you treat the bacterial organisms?

- 1) Yes
- 2) No
- 3) Maybe – it depends

Talk goals

- Explain how mNGS fits into the infectious disease diagnostic armamentarium
- Review evidence for plasma mNGS for diagnosis of immunocompromised patients with pulmonary lesions
- Review evidence for plasma mNGS for diagnosis in neutropenic fever
- Discuss the current UCSF guidelines for plasma mNGS use

Metagenomic sequencing



Go Fish and diagnostic bias

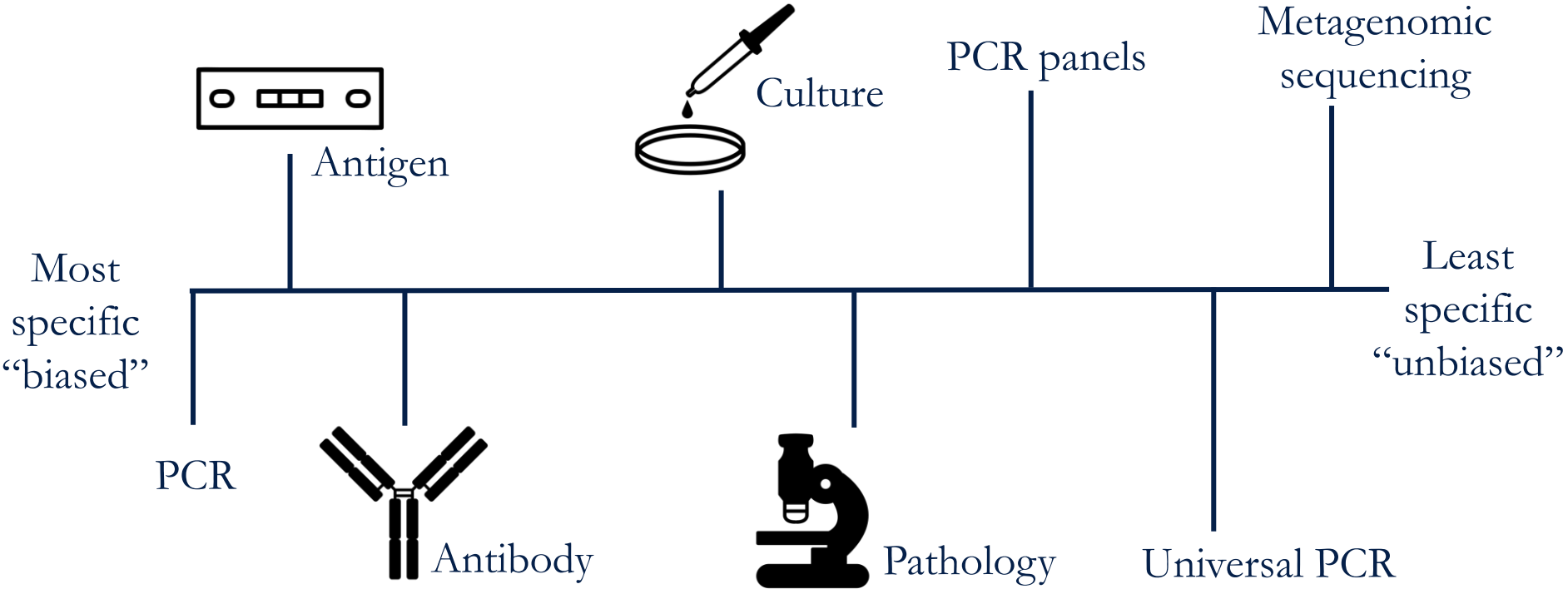
A biased test answers the question: “Do you have a *yellow* fish?”

An unbiased test answers the question: “What fish do you have?”

Most tests are *biased*



Clinical diagnostics summary



A clinician's view of mNGS challenges

Specificity

Technical contaminants

Delftia, other water-dwelling organisms

Biologically present but
of unclear relevance

Frequent detection of GI/oral flora in critically ill patients.
mNGS plasma detection \neq blood culture.
DNA viruses of unclear significance
Risk of **inappropriate antimicrobial escalation**

Sensitivity

Negative predictive value
not known

Most likely not as sensitive as PCR given no amplification steps
Unclear how to interpret a negative
Risk of **inappropriate antimicrobial de-escalation**

What makes a mNGS diagnosis high impact?

Pathogen

Enriched for bugs that don't culture

Enriched for "never commensals"

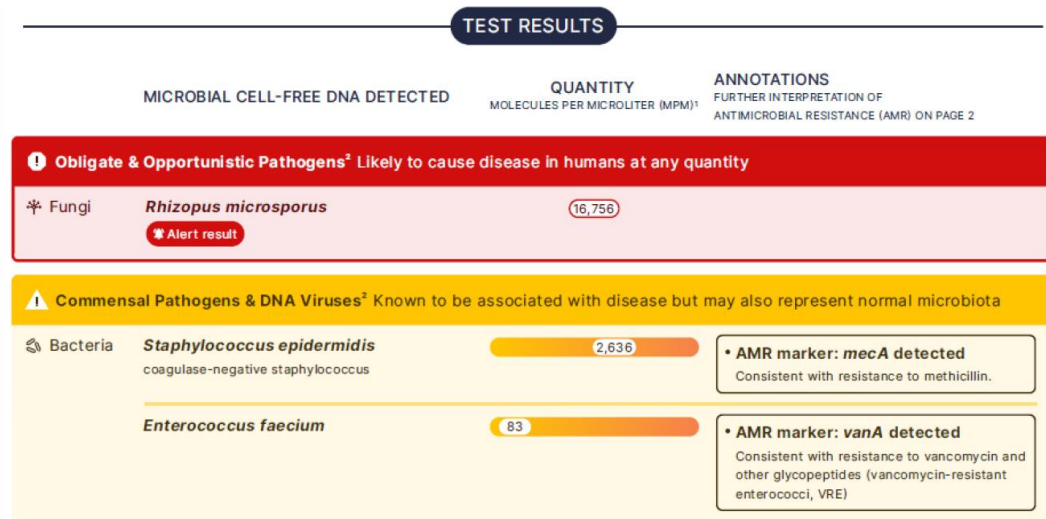
Change management (appropriately)

Patient

Immunocompromised patients at risk of wide pathogen variety

Life-threatening conditions

Interpreting plasma mNGS: the bugs



¹ Molecules Per Microliter = number of DNA fragments present in one microliter of plasma. Visualization of MPM shows quantile of each detected microbe based on 10,000 specimens with positive, quantitative Karius Test results. No quantile is shown if < 20 detections of the microbe were made in the 10,000 specimens or if the microbe is an obligate or opportunistic pathogen. The analytical range of the assay is 10 - 316,000 MPM.

² Based on a review of Carroll KC, Pfaller MA. 2019. Manual of Clinical Microbiology, 12th Edition. ASM Press, Washington, DC and Bennett JE, Dolin R, Blaser MJ. 2019. Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases, 9th Edition. Elsevier, Philadelphia, PA

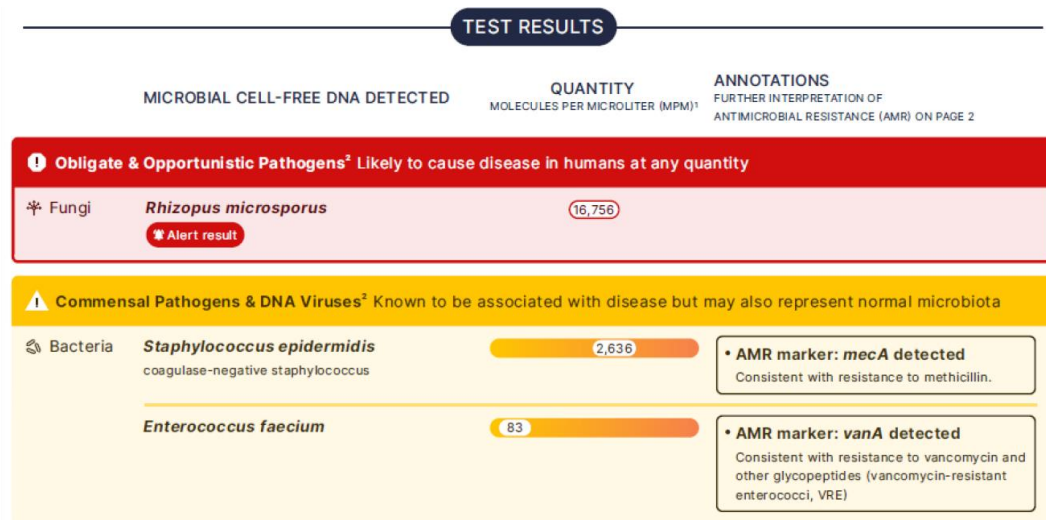
Obligate/Opportunistic pathogens

- Always assume true pathogen = most fungi, highly pathogenic bacteria (*Legionella*)

Commensals/DNA Viruses

- Do not assume that mNGS detection = bacteremia
- May indicate gut or mouth “leak”
- DNA viruses may be present but not causing pathology
- We send orthogonal test if unclear (CMV PCR, bacterial blood culture)

Interpreting plasma mNGS: MPM and AMR




¹ Molecules Per Microliter = number of DNA fragments present in one microliter of plasma. Visualization of MPM shows quantile of each detected microbe based on 10,000 specimens with positive, quantitative Karius Test results. No quantile is shown if < 20 detections of the microbe were made in the 10,000 specimens or if the microbe is an obligate or opportunistic pathogen. The analytical range of the assay is 10 - 316,000 MPM.

² Based on a review of Carroll KC, Pfaller MA. 2019. Manual of Clinical Microbiology, 12th Edition. ASM Press, Washington, DC and Bennett JE, Dolin R, Blaser MJ. 2019. Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases, 9th Edition. Elsevier, Philadelphia, PA

- MPM = molecules per microliter, rough estimate of how much pathogen is present
- May be correlated with PCR measures within a particular organism (e.g. CMV)
- Cannot really compare across different organisms
- Antimicrobial resistance (AMR) is in beta-testing. Our practice:
 - Gram-positives: better data, may broaden abx based on this result
 - Gram negatives: really insufficient data, would not change therapy

Plasma mNGS: the logistics

- Goes to commercial company (Karius) 
- Turn around time variable, but usually 3-4 weekdays
- Tests are often rejected due to incorrect collection
- Cost: ~\$1500/sample

Talk goals

- Explain how mNGS fits into the infectious disease diagnostic armamentarium
- Review evidence for plasma mNGS for diagnosis of immunocompromised patients with pulmonary lesions
- Review evidence for plasma mNGS for diagnosis in neutropenic fever
- Discuss the current UCSF guidelines for plasma mNGS use

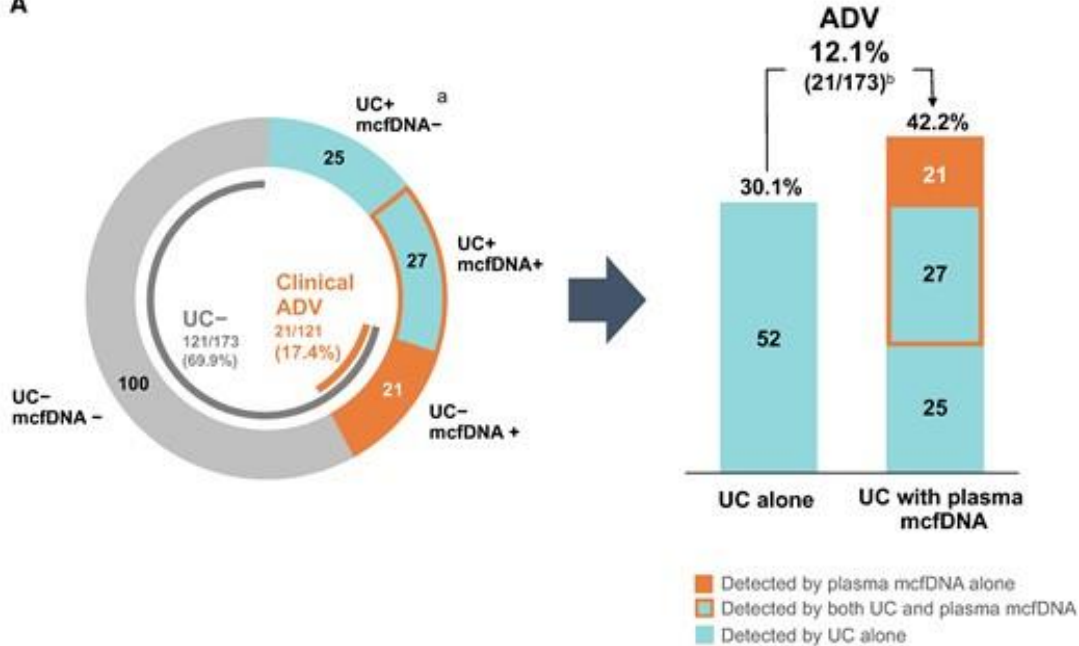
Severely immunocompromised patients with pneumonia

- Key study: PICKUP
 - Multicenter, prospective, observational study comparing Karius mNGS to standard of care in immunocompromised adults*
 - All patients had pneumonia of unclear etiology



How Karius performed in PICKUP

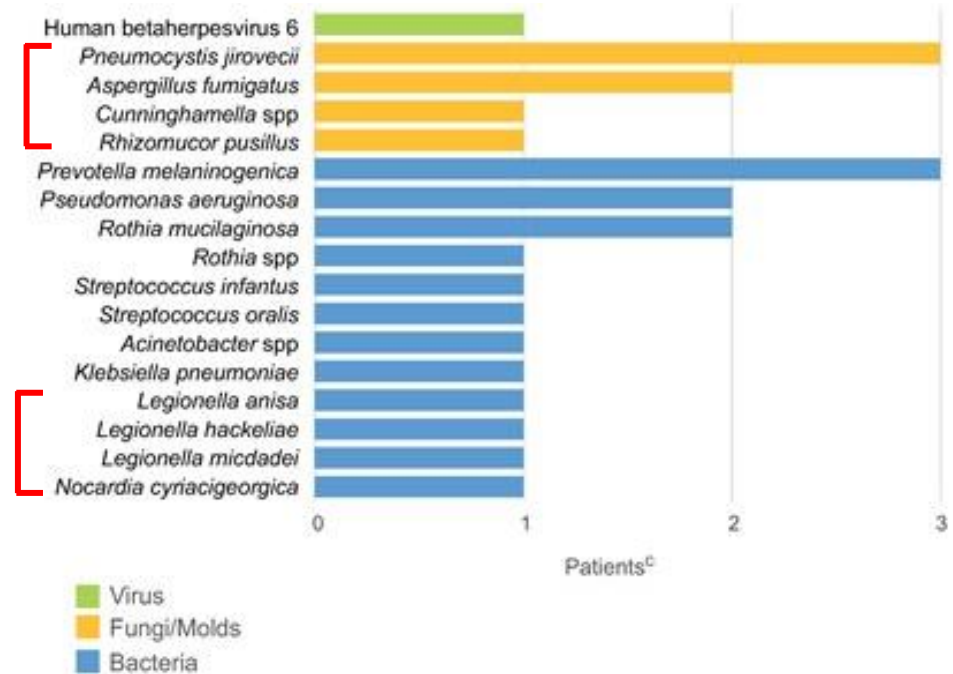
A



- 30.1% of patients received a diagnosis through usual care (UC)
- 42.2% of patients received a diagnosis through a composite of UC and Karius testing
- 21/173 (12.1%) patients exclusively diagnosed by Karius

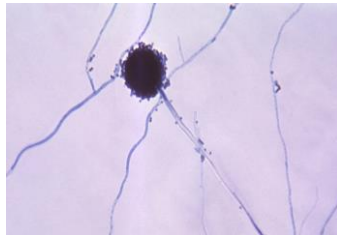
Patients diagnosed only by mNGS

- Of the 21 patients exclusively diagnosed by Karius, pathogens included fungi, bacteria, and viruses
- Retrospective review indicated that 17 diagnoses might have changed management (9.8% total patients)
- Notably, not all patients can get bronchoscopies, so this may *underestimate* benefit



The diagnoses missed by mNGS

- 25/173 patients (14.5%) had positive diagnoses by usual care but negative Karius tests
- Inherently misses RNA viral pathogens
- Intriguing finding that mNGS may be worse at detection of *Aspergillus* than other fungi
- Another study (Hill et al 2021) have also shown what appears to be worse detection of *Aspergillus*



Bacteria N = 7

- All common respiratory bacterial pathogens

Viruses N = 4

- All RNA viruses
- Rhinovirus, SARS-CoV-2, parainfluenza

Fungi N = 17

- 1 *Cryptococcus*
- 16 *Aspergillus* species

Talk goals

- Explain how mNGS fits into the infectious disease diagnostic armamentarium
- Review evidence for plasma mNGS for diagnosis of immunocompromised patients with pulmonary lesions
- Review evidence for plasma mNGS for diagnosis in neutropenic fever
- Discuss the current UCSF guidelines for plasma mNGS use

Defining the problem of FN

Concern for infection

Core temperature

≥ 38.3 C once or
 ≥ 38.0 C for ≥ 1
hour



Profound immunocompromise

UCSF definition:

Absolute neutrophil count
 < 500 *or* < 1000 and falling

Usually from:

Cancer

Chemotherapy

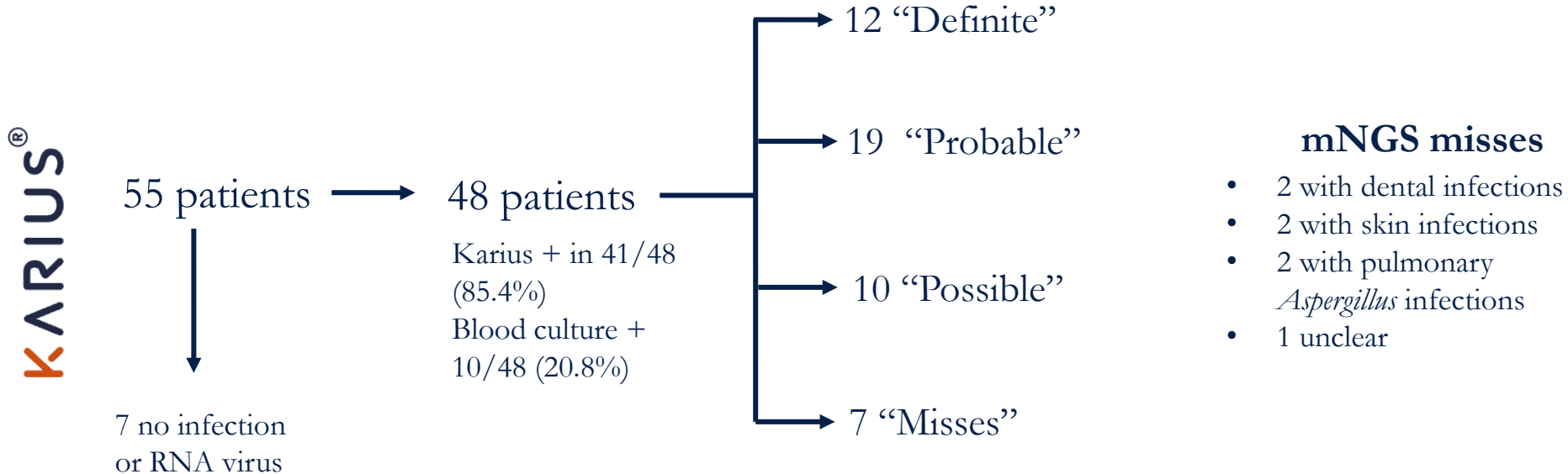
Bone marrow transplant

- Most patients are on prophylactic antibiotics if neutropenic, before developing fever
- Only 20-30% of patients usually receive a microbiologic diagnosis

A key febrile neutropenia study



mNGS and neutropenic fever: the results



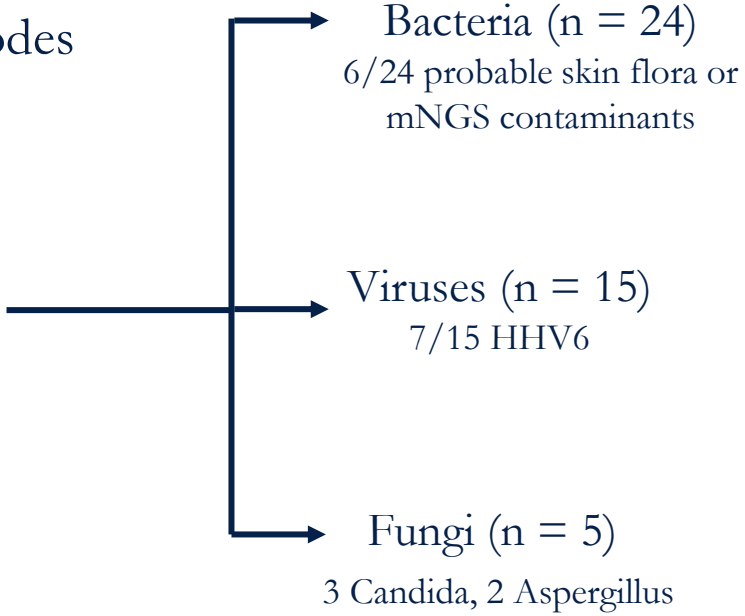
Authors estimate antimicrobial management would have changed in 26 patients (47.3%), but this assumes de-escalation on negative results...

A second FN study highlights hurdles

DISQVER[®]

98 FN episodes
61 patients

DISQVER + in
42/97 (43.3%)
Blood culture +
14 (14.3%)



Clinical Utility

In patients with +mNGS, ID
doctors tried to answer:
Would this result have
changed management?

**In 60% of cases, doctors
could not agree**

Schulz et al. Open Forum Infectious Disease. 2022.

Talk goals

- Explain how mNGS fits into the infectious disease diagnostic armamentarium
- Review evidence for plasma mNGS for diagnosis of immunocompromised patients with pulmonary lesions
- Review evidence for plasma mNGS for diagnosis in neutropenic fever
- Discuss the current UCSF guidelines for plasma mNGS use

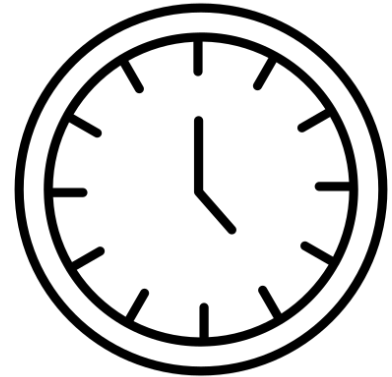
Question 3: How often do you use metagenomic next-generation sequencing?

- 1) Never
- 2) 1-2x a year
- 3) Monthly
- 4) Weekly
- 5) Daily

Defining timing

- **First line** = Sent concurrently with other immediate diagnostics (blood cultures, imaging, etc).
 - For inpatients, implies from the ED or immediately after admission.

- **Second line** = Sent following other syndrome-appropriate first-line diagnostics, including blood cultures, cryptococcal antigen, viral PCRs, and/or imaging.
 - For inpatients, implies ~2-3 days after admission.



UCSF Plasma mNGS: First Line Indications

1. Severely immunocompromised* patient with pneumonia, especially if:

- Concern for high concern for atypical infection such as IFI, Nocardia, Legionella
- Not responding to standard care

3. Fulminant infection *and* strong epidemiologic concern for atypical infection:

- High specific concern for disseminated *M. tuberculosis*, Nocardia, Coxiella/Q fever, Brucella, tularemia; invasive fungal infection

2. Fulminant CNS infection *and* sampling of CSF/CNS is not feasible or delayed

- High concern for CNS infection that reflects disseminated illness
- CNS/CSF sampling not technically possible

*Severely immunocompromised = bone marrow transplant within 1 year, solid organ transplant within 1 year, primary severe immunodeficiency

UCSF Plasma mNGS: Second Line Indications

1. Culture negative endocarditis *and*

- Negative first line workup (blood cultures, serological tests/antibodies as indicated)

3. Deep seated lesions/abscesses (epidural, hepatic, splenic, peri-renal, pleural effusion) *and*

- Blood cultures negative
- Sampling of site unfeasible and/or unrevealing

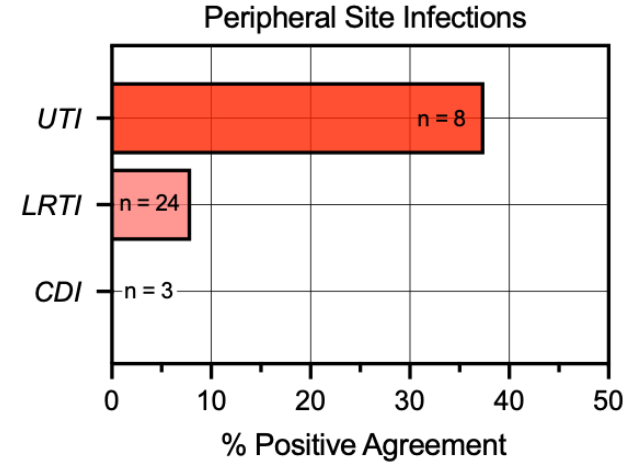
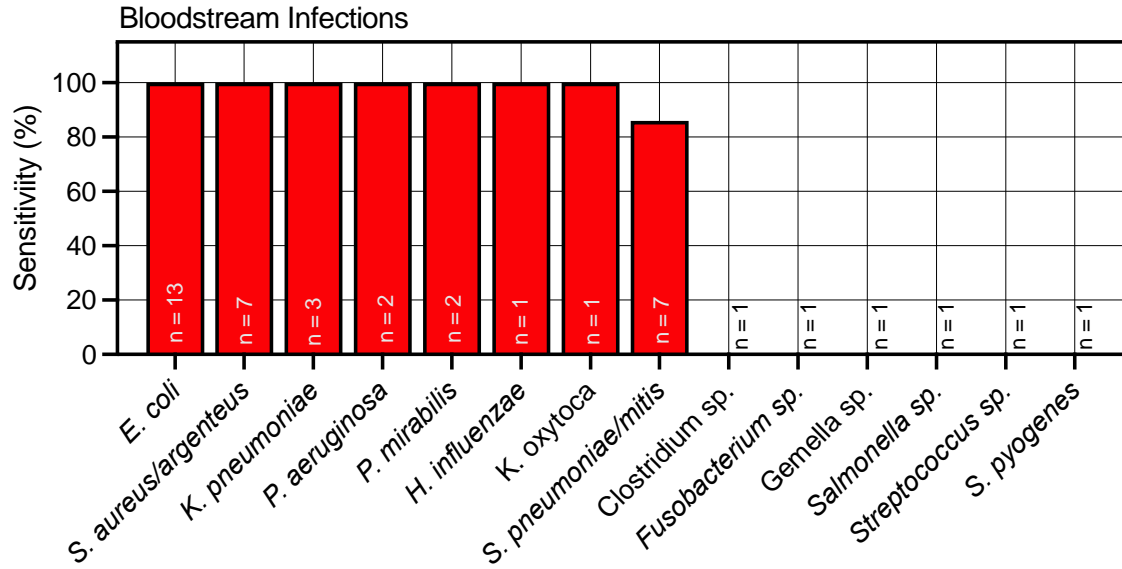
2. Fever of unknown origin *and*

- Negative first line workup (blood cultures, imaging, malignancy/autoimmune workup)
- If patient is on empiric antibiotics, a trial of stopping them should be considered

4. Persistent febrile neutropenia *and*

- Negative first line workup (blood cultures, cross-sectional imaging)

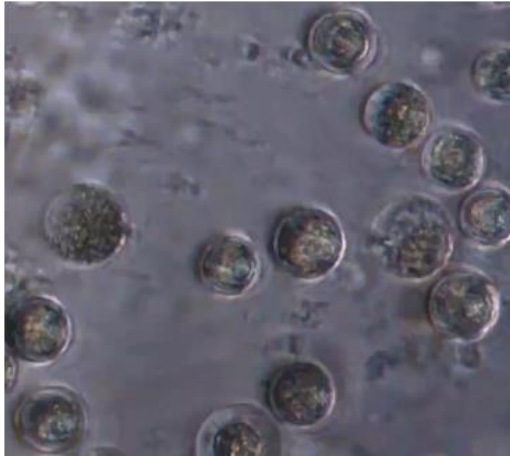
Emerging indications: could mNGS be useful in sepsis?



- Specificity 78%
- Possible pathogens identified in 42% of patients with culture negative sepsis

Plasma mNGS can sometimes diagnose CNS infection

- Patient with no past medical history developed neurological symptoms and fever.
- Lumbar puncture consistent with infection, but CSF cultures negative.
- Imaging ambiguous: infection or non-infectious autoimmune process
- Plasma mNGS sent (Karius). Patient discharged



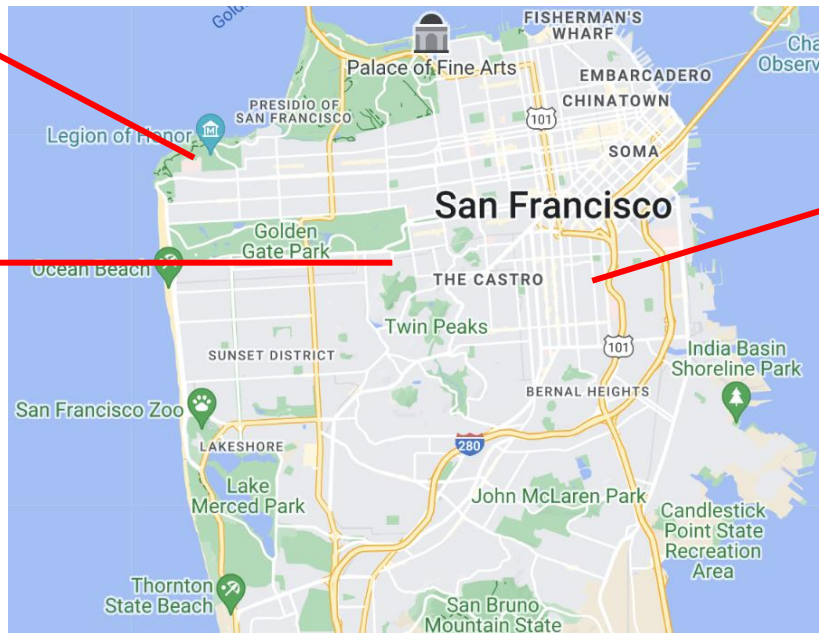
- **Diagnosis: *Balamuthia mandrillaris*.**
- Multidisciplinary ameba team promptly involved
- Therapy started within 1-2 days

A word on equity: a tale of three hospitals

San Francisco
Veterans
Association

UCSF
Parnassus

Zuckerberg San
Francisco General
Hospital



Conclusions

- Metagenomic sequencing is an exciting new tool to understand what is making patients sick
- Uniquely can detect pathogens that the physician may not be thinking about (unbiased diagnostic).
- Benefit of plasma mNGS is dependent on syndrome being tested
- Perhaps the strongest evidence is for highly immunocompromised patients with pneumonia
- More work is needed to determine the best use case of this test!

Thank you! Questions?



References

1. R. B. Hill, Jr., D. T. Rowlands, Jr., D. Rifkind, Infectious pulmonary disease in patients receiving immunosuppressive therapy for organ transplantation. 1964. *Rev Med Virol* 9, 5-10; discussion 13-15, 10-14 (1999).
2. S. P. Bergin *et al.*, Plasma Microbial Cell-Free DNA Sequencing in Immunocompromised Patients with Pneumonia: A Prospective Observational Study. *Clin Infect Dis*, (2023).
3. J. A. Hill *et al.*, Liquid Biopsy for Invasive Mold Infections in Hematopoietic Cell Transplant Recipients With Pneumonia Through Next-Generation Sequencing of Microbial Cell-Free DNA in Plasma. *Clin Infect Dis* 73, e3876-e3883 (2021).
4. A. G. Freifeld *et al.*, Clinical practice guideline for the use of antimicrobial agents in neutropenic patients with cancer: 2010 update by the infectious diseases society of america. *Clin Infect Dis* 52, e56-93 (2011).
5. E. Benamu *et al.*, Plasma Microbial Cell-free DNA Next-generation Sequencing in the Diagnosis and Management of Febrile Neutropenia. *Clin Infect Dis* 74, 1659-1668 (2022).
6. E. Schulz *et al.*, Pathogen Detection by Metagenomic Next-Generation Sequencing During Neutropenic Fever in Patients With Hematological Malignancies. *Open Forum Infect Dis* 9, ofac393 (2022).
7. L. M. Baddour *et al.*, Infective Endocarditis in Adults: Diagnosis, Antimicrobial Therapy, and Management of Complications: A Scientific Statement for Healthcare Professionals From the American Heart Association. *Circulation* 132, 1435-1486 (2015).
8. E. M. Eichenberger *et al.*, Microbial Cell-Free DNA Identifies Etiology of Bloodstream Infections, Persists Longer Than Conventional Blood Cultures, and Its Duration of Detection Is Associated With Metastatic Infection in Patients With Staphylococcus aureus and Gram-Negative Bacteremia. *Clin Infect Dis* 74, 2020-2027 (2022).
9. N. Degner, Smollin, M., Equils, O., Arun, A, DeVries, C., MacIntyre, A., Ahmed, A.A., Rapid, Non-invasive Detection of Invasive Bartonella Infections in Pediatric Patients Using the Karius Test, a Next-Generation Sequencing Test for Microbial Cell-free DNA in Plasma. *Journal of the Pediatric Infectious Disease Society* 10, S11 (2021).
10. K. L. Kalantar *et al.*, Integrated host-microbe plasma metagenomics for sepsis diagnosis in a prospective cohort of critically ill adults. *Nat Microbiol*, (2022).
11. D. Ren *et al.*, The microbiological diagnostic performance of metagenomic next-generation sequencing in patients with sepsis. *BMC Infect Dis* 21, 1257 (2021).
12. M. R. Wilson *et al.*, Clinical Metagenomic Sequencing for Diagnosis of Meningitis and Encephalitis. *N Engl J Med* 380, 2327-2340 (2019).
13. N. Spottiswoode *et al.*, Successful Treatment of Balamuthia mandrillaris Granulomatous Amebic Encephalitis with Nitroxoline. *Emerg Infect Dis* 29, 197-201 (2023).
14. A. D. Kerkhoff, R. L. Rutishauser, S. Miller, J. M. Babik, Clinical Utility of Universal Broad-Range Polymerase Chain Reaction Amplicon Sequencing for Pathogen Identification: A Retrospective Cohort Study. *Clin Infect Dis* 71, 1554-1557 (2020).
15. A. D. Kerkhoff, Matzko M., Chiu C., Miller S., Babik J. M., The Comparative Utility Of Metagenomic Next-Generation Sequencing and Universal PCR for Pathogen Detection on Cerebrospinal Fluid: A Retrospective Analysis From a Tertiary Care Center. *Open Forum Infect Dis* 6, S134-135 (2019).
16. C. R. Langelier *et al.* Integrating host response and unbiased microbe detection for lower respiratory tract infection diagnosis in critically ill adults. *PNAS*. 115 (52): E12353-E12362